Review

Role of ferroptosis in nanofiber-induced carcinogenesis

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Abstract

Biopersistent nanofibers with specified physical dimension are unexpected human carcinogens whether they are natural or synthetic. Asbestos, a natural fibrous mineral, is classified as a definite human carcinogen (IARC Group 1) to cause malignant mesothelioma (MM) and lung cancer. Multi-walled carbon nanotube of 50 nm-diameter was defined in 2014 as a possible carcinogen (IARC Group 2B) toward MM, fortunately with no authorized patients thus far. Carcinogenic mechanism of asbestos has been a mystery for a long time. It is now recognized that asbestos goes through lung parenchyma by collecting hemoglobin-derived iron to reach pleural cavity, which takes several decades. Iron-loaded asbestos can induce oxidative damage directly to mesothelial cells, carcinogenesis-target cells lining somatic cavities. Recently, it was clarified that surrounding stromal environment are as important for mesothelial carcinogenesis. The novel concept here is ceaseless ferroptosis of macrophages, which forms a Fe(II)-dependent stromal mutagenic milieu indirectly for mesothelial cells and indeed is a revised understanding of frustrated phagocytosis. Deposition of foreign materials eventually causes iron accumulation *in situ* due to the innate characteristic of preserving iron inside cells. Nanofiber-induced carcinogenesis may be involved in other human carcinogenesis, including ovarian cancer. Alternatively, iron excess can be an optimal target of cancer prevention and cancer treatment.

Key words: asbestos, mesothelioma, ferroptosis, macrophage, iron, carbon nanotube

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Abbreviations:

DMT1	divalent metal transporter 1		
EV(s)	extracellular vesicle(s)		
IARC	International Agency for Research on Cancer		
IRE	iron-responsive element		
IRP(s)	iron-regulatory protein(s)		
MM	malignant mesothelioma		
MWCNT	multi-walled carbon nanotube		
NCOA4	4 nuclear receptor coactivator 4		
PCBP1/2	1/2 poly-repeated cytidine-binding protein 1/2		
UTR	untranslated region		



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Introduction

Cancer is one of the leading causes of human mortality all over the world (https://www.who.int/data/gho/data/themes/ mortality-and-global-health-estimates). Whereas molecular carcinogenic mechanism of each human cancer is diversely different and unidentified in most of the cases [1], some of the environmental carcinogenesis reveals unequivocally significant epidemiological association, including ionizing radiation with leukemia [2] and asbestos exposure with malignant mesothelioma (MM) [3].

Asbestos is a natural fibrous mineral, which has been used in the human history since 2,500 B.C. due to its resistance to heat, acid and friction, such as in pottery and sacred cremation garments for the Egyptian pharaohs [4]. After the industrial revolution period, wide use of asbestos started worldwide because of the economical merits of mining [5], which continued till epidemiologist recognized the association between asbestos exposure and MM or lung cancer [6]. MM has been and is a rare tumor [7] in that the tumor retains the characteristics of thin and flat mesothelial cells which line somatic cavities and decrease the friction-derived heat by producing hyaluronic acid [8]. We have ~2,000 new patients yearly in Japan (https://www.mhlw.go.jp/toukei/saikin/hw/jinkou/tokusyu/chuuhisyu17/dl/chuuhisyu.pdf) in comparison to ~120,000 new lung cancer patients. Historically, prolonged average human life was essential for the recognition of this tumor due to an extremely long incubation period of 30~40 years after asbestos exposure, which has been a long-time mystery [9]. After the recognition of all the asbestos as a definite human

carcinogen (Group 1) by the International Agency for Research on Cancer (IARC) in 1987 [5], scientists have lost interest in the mechanistic elucidation for a while. Of note, asbestos is negative for Ames test [10].

In Japan, asbestos issue was reminded in 2005 when Kubota shock occurred by newspaper report [11], when inhabitants near the asbestos factory obtained a high incidence of MM. Even now the prognosis of MM is quite poor because of the difficulty in diagnosing the early stage of MM [12]. It is established that carcinogenicity of asbestos fibers depends on its high affinity for histones and hemoglobin-derived iron. Indeed, asbestos (ferruginous) body found in the lung parenchyma of those people exposed to high amounts of asbestos supports this mechanism, thus generating physical scissors for cutting genomic DNA via the Fenton reaction [3, 9] (Figure 1). Size $(3 \text{ nm} \sim 5 \mu \text{m in diameter})$ [13] and length as an aspect ratio (fiber length/ diameter) of > 3 were important to reach alveolar space and this needle-like structure with biopersistence was essential to reach pleural and sometimes peritoneal cavities eventually.

Asbestos Chest wall Direct effect Pleural cavity DNA Doublestrand Break DNA Doublestrand Break DNA Doublestrand Break Asbestos surface DNA Doublestrand Break Machine Strangen Stran

Figure 1. Current understanding of the mechanism of asbestos-induced mesothelial carcinogenesis in humans.

Note that a few decades are required for asbestos fibers to go through lung parenchyma to the pleural cavity. Asbestos fibers have hemolytic activity and high affinity for hemoglobin, thus accumulating massive amounts of iron on its surface (red fibers in the figure) to cause DNA double-strand breaks via Fenton reaction in parietal mesothelial cells, targets for carcinogenesis (direct effect). Refer to text for details.

Iron metabolism revised

No life on the earth can live without iron from bacteria to humans [14, 15]. There are recent advancements in the understanding of iron metabolism in higher animals, which started from transferrin/ferritin system, iron transporters (DMT1, ferroportin, etc.) [16], posttranscriptional regulation (IRE-IRPs system) and IRP2/FBXL5-ubiquitin-proteasome system [17, 18].

The recent noteworthy new concepts in iron metabolism would be ferritinophagy [19, 20], cytosolic iron chaperones [21] and ferritin secretion via IRE-IRP/CD63-regualted extracellular vesicles [22]. All of these suggest that intracellular iron levels are strictly regulated not to abandon but efficiently reuse iron and to maintain the iron in a safe non-catalytic fashion. Ferritinophagy

is ferritin-specific autophagic process directed by nuclear receptor coactivator 4 (NCOA4) to release iron from ferritin cores into lysosomes [23]. Poly(rC)-binding protein 1 and 2 (PCBP1/2) has been first reported as intranuclear RNA-binding proteins but are now recognized as mutually exclusive cytosolic iron chaperones [24-26]. Only PCBP1 can load Fe(II) to ferritin cores whereas PCBP2 play a role in other intracellular Fe(II) deliveries [27, 28]. Theoretically, PCBP1/2 carries 3 molecule of Fe(II) in non-catalytic manner [21]. In general, PCBP1 works as tumor suppressor gene [29] and PCBP2 as oncogene [30]. The last one is quite new reported in 2021. We discovered a canonical IRE in the 5'-untranslated region (UTR) of CD63 mRNA responsible for regulating its expression in response to increased iron. We showed that under iron-loading, intracellular ferritin is transferred via NCOA4 to CD63(+) extracellular vesicles (EVs) that are then secreted. Such iron-dependent secretion of the major iron storage protein ferritin is performed through CD63(+) EVs [22].

Ferroptosis

Ferroptosis is a recently defined regulated necrosis. The characteristic in this cell death mode is the dependence on catalytic Fe(II) leading to lipid peroxidation [18, 31]. This was first reported on *H-ras* mutated fibrosarcoma cells with the use of erastin, an inhibitor for cystine/glutamate antiporter (SLC7A11), resulting in decrease in reduced GSH as an antioxidant [32]. The current revised concept of ferroptosis is the imbalance between Fe and S (-SH; sulfhydryls) in favor of iron, causing Fenton-reaction [18]. Iron is one of the most basic elements of the cell, working as cofactors in enzymes, such as ribonucleotide reductase (DNA synthesis), cytochrome oxidase (energy production) and catalase (antioxidant), and hemoglobin in higher species. Thus, every cell tries to maintain the amounts of iron, and bacterial and fungal infections might be a fight to obtain iron for the continued growth of those invaders [15].

It is worth mentioning here that there is no mechanisms to abandon iron from an individual in higher species except for bleeding though Fe(II) can be secreted extracellularly via ferroportin [25, 33]. Accordingly, accumulated iron or decreased antioxidant systems results in ferroptosis. Cancer cells collects iron for persistent proliferation [34-36]. Thus, it is not hard to imagine that cancer cells specifically fall into ferroptosis when the overused antioxidant pathway is squeezed with certain chemicals [1]. Furthermore, autophagic process promotes ferroptosis via ferritin degradation [37] whereas ferritinophagy inhibition via NCOA4 deficiency in the heart mitigates the development of pressure overload-induced dilated cardiomyopathy [38].

Novel mechanism in asbestos-induced mesothelial carcinogenesis

Mesothelial cells are the major target cells in asbestos-induced carcinogenesis. Thus, the research has been performed to clarify how asbestos causes genotoxicity on mesothelial cells directly. However, we recently recognized that the surrounding microenvironment is as important for mesothelial carcinogenesis, based on animal model experiments [39]. In this peritoneal injection model in rats, we noticed after 1 month of injection that virtually all the asbestos fibers are inside the macrophages, which formed granuloma, a foreign body reaction.

Granuloma formation is a specific inflammation mainly of macrophages to confine uncontrollable materials/agents within those barriers, whether they are independent species or man-made synthetic materials (**Figure 2**). Macrophages are phagocytic and antigen-presenting cells. Furthermore, they play a central role in iron metabolism, especially regarding iron recovery from dead or dying cells. We found that asbestos due to its specified physical dimension kills macrophages consistently at first via lysosomal-dependent cell death and finally ferroptosis, which generates Fe(II)-dependent mutagenic stromal milieu, causing β -catenin induction in mesothelial cells [39]. This is an indirect effect to mesothelial cells and may be a revised understanding of frustrated phagocytosis [40].

Carbon nanotubes

Carbon nanotube was discovered with electron microscopic observation in 1991 [41]. This is a synthetic material, consisting exclusively of carbon and with a tubular structure of a few nanometer (single-walled) to several hundred nanometers (multi-walled) in diameter. This material has been and is used for numerous industrial purposes, such as in lithium battery and liquid crystal film, based on its physical nature of rigidity, electrophilicity and heat conductivity [42, 43].

However, its similarity to asbestos fibers in physical dimension was questioned in the early 2,000's, and thus we worked on this issue. We found that diameter of carbon nanotube is the most critical risk factor, where multi-walled carbon nanotube (MWCNT)



Figure 2.Stromal mutagenic milieu generated by ceaseless
macrophage ferroptosis.

Mesothelial cells have phagocytic activity and asbestos fibers in the somatic cavity are eventually transferred to stromal tissue supporting the somatic walls, where macrophages take up the fibers coated with hemoglobin-derived iron. Macrophages try to accommodate all the asbestos fibers by making granuloma, a collection of macrophages with multinucleated giant cells. However, many of the macrophages die through ferroptosis because they cannot cope with the disposal of these thin and long fibers. Finally, abundant iron is released to the stroma of somatic wall, which constitutes the mutagenic milieu for the surface-lining mesothelial cells (indirect effect).

of ~50 nm-diameter was most carcinogenic to mesothelial cells in rat intraperitoneal injection studies [44] and that MWCNT of ~15 nm-diameter was not carcinogenic to mesothelial cells even after > 3 years of observation [45]. The interesting point was that carbon nanotubes have a high affinity not only for histone and hemoglobin but also for transferrin, an iron transporting protein in the serum, and that only MWCNT of -50 nm could go into mesothelial cells [46]. There observations suggest that excess iron play a role in MWCNT-associated mesothelial carcinogenesis. This is strongly confirmed with the similar genetic alterations observed in the rat and human MMs between asbestos origin [47-49] and MWCNT of 50 nm-diameter origin [44], where homozygous deletion of $p16^{lnk4a}$ tumor suppressor gene was the most prominent . Indeed, we believe that deletion of $p16^{lnk4a}$ tumor suppressor gene is a marker of Fenton reaction-induced carcinogenesis [35, 50], based on the studies of ferric nitrilotriacetate (Fe-NTA)-induced renal carcinogenesis model (**Table 1**). Regarding Fe-NTA-induced renal carcinogenesis,

Table 1.	Similarities and differences among	a three <i>wild-typ</i>	e animal models ca	using cancer throug	h excess iron.
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Models	Fe-NTA	Asbestos (chrysotile, crocidolite and amosite)	MWCNT of 50-nm diameter
Species	Rat, mouse	Rat, mouse	Rat, mouse
Injection	<i>ip</i> , 3~5 times a week/10-12 weeks	<i>ip</i> , 1~3 times	<i>ip</i> , 1~3 times
Major pathology	Repeated Fenton reaction in the renal proximal tubules	Direct action to mesothelial cells with prolonged foreign body reaction (ceaseless ferroptosis of macrophages)	Direct action to mesothelial cells with prolonged foreign body reaction
Origin of excess iron	Fe-NTA itself	Asbestos itself (crodidolite, amosite), hemolysis (chrysotile) and affinity to hemoglobin	Affinity to hemoglobin and transferrin
Induced cancer	Renal cell carcinoma; 50% of pulmonary metastasis in rats	Malignant mesothelioma	Malignant mesothelioma
Major target gene	Homozygous deletion of <i>16^{ink4a}</i> tumor suppressor gene	Homozygous deletion of <i>16^{ink4a}</i> tumor suppressor gene	Homozygous deletion of 16 ^{ink4a} tumor suppressor gene
References	[15, 50, 71, 72]	[49, 73]	[44, 46]

refer to other published reviews [15, 51].

Recently, Tim4 was identified as a receptor for MWCNT in macrophages leading to granuloma formation [52, 53]. It was recently reported that Tim4+ macrophages sequester and impair proliferation of CD8+ T cells [54] whereas TIM4 expression by dendritic cells mediates uptake of tumorassociated antigens and anti-tumor responses [55]. Whether persistent ferroptosis of macrophages is at work for mesothelial carcinogenesis requires further investigation. We have to stress here that no definite MM case has been reported thus far in terms of carbon nanotubes. We believe that this depends on early recognition (IARC Group 2B) [56] and the efforts on the industry side to avoid the use of high-risk carbon nanotubes and to develop the large-scale automated systems to minimize human exposure. The continued use of MWCNT in industry and in material science, even for the robotic systems [57], is very different from the asbestos case and we can declare that this is one of the successful examples of experimental pathology using animal models and regulatory science.

Ovarian carcinogenesis

Another example we suspect for nanofiber-induced carcinogenesis accompanied by excess iron is ovarian carcinogenesis. Endometriosis is a female disease of reproductive



Toyokuni S. et al.





Asbestos and other nanofibers may exist in the vulva of reproductive-age women as contaminants of family laundry or cosmetic applications. Liquid flow from vulva, vagina, uterine cavity, oviduct to ovary may carry these fibers to ovarian epithelial cells. Refer to text for details.

age where endometrium exists outside of uterine cavity, leading to iron excess due to monthly bleeding. In the case of ovarian endometriosis, this iron excess is established as a risk for adenocarcinoma, especially clear cell carcinoma and endometrioid adenocarcinoma, in addition to menstrual pain and infertility [58, 59]. In addition to this, fibrous materials, including asbestos, are suspected to be carcinogenic to ovarian epithelial cells. There are two points that are true but not well recognized: 1) there is a pathway in women through vagina, endometrium, oviduct to ovary [60-62], which happens at fertilization; 2) frequent use of baby power, including talc and some amounts of asbestos, may be epidemiologically associated with ovarian cancer [63, 64] though it is still controversial [65, 66] (Figure 3). We believe that this is an important question to be explored to prevent ovarian carcinogenesis.

Conclusion

Biopersistent fibrous materials may provide humans with unexpected risk of cancer, depending on its physical parameters and exposure route. This is closely associated with the general response of our cells against foreign materials to collect/recover iron as much as possible and to deplete iron in the extracellular space. Alternatively, if the macrophages cannot accommodate or scavenge these fibrous materials and die through ferroptosis, mutagenic stromal milieu is generated [39]. Finally, it is important to recognize that iron excess can be a target for cancer prevention [67, 68] and also for cancer therapy [18, 69, 70], including non-thermal plasma.

Declaration of competing interest

The authors declare no conflict of interest to present.

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References

- [1] Toyokuni S, Ito F, Yamashita K, Okazaki Y, Akatsuka S: Iron and thiol redox signaling in cancer: An exquisite balance to escape ferroptosis. Free Radic Biol Med 108: 610-626, 2017.
- [2] Leuraud K, Richardson DB, Cardis E, Daniels RD, Gillies M, et al.: Risk of cancer associated with low-dose radiation exposure: comparison of results between the INWORKS nuclear workers study and the A-bomb survivors study. Radiat Environ Biophys 60: 23-39, 2021.
- [3] Toyokuni S: Iron addiction with ferroptosis-resistance in asbestos-induced mesothelial carcinogenesis: Toward the era of mesothelioma prevention. Free Radic Biol Med 133: 206-215, 2019.
- [4] Oury TD, Sporn TA, Roggli VL (eds): Pathology of Asbestos-Associated Diseases, Third edition, Springer, Berlin/Heidelberg, 2014.
- [5] IARC, WHO, Asbestos (chrysotile, amosite, crocidolite, tremolite, actinolite, and anthophyllite), IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. A Review of Human Carcinogens; Part C: Arsenic, Metals, Fibres, and Dusts, Lyon, France, 2012, pp. 219-309.
- [6] Wagner JC, Sleggs CA, Marchand P: Diffuse pleural mesothelioma and asbestos exposure in North Western Cape Province. Br J Ind Med 17: 260-271, 1960.
- [7] Alpert N, van Gerwen M, Taioli E: Epidemiology of mesothelioma in the 21st century in Europe and the United States, 40 years after restricted/banned asbestos use. Transl Lung Cancer Res 9: S28, 2020.
- [8] Husain AN, Colby T, Ordonez N, Krausz T, Attanoos R, et al.: Guidelines for pathologic diagnosis of malignant mesothelioma: 2012 update of the consensus statement from the International Mesothelioma Interest Group. Arch Pathol Lab Med 137: 647-667, 2012.
- [9] Toyokuni S: Mechanisms of asbestos-induced carcinogenesis. Nagoya J Med Sci 71: 1-10, 2009.
- [10] Clift MJD, Raemy DO, Endes C, Ali Z, Lehmann AD, et al.: Can the Ames test provide an insight into nano-object mutagenicity? Investigating the interaction between nano-objects and bacteria. Nanotoxicology 7: 1373-1385, 2013.
- [11] 79 Kubota workers killed by asbestos over 26 years (2005.06.30), The Daily Yomiuri, 2005.
- [12] Asciak R, George V, Rahman NM: Update on biology and management of mesothelioma. Eur Respir Rev 30: 200226, 2021.
- [13] Witschi HR, Last JA: Toxic responses of the respiratory system. Casarett and Doull's toxicology: the basic science of poisons. 6th ed. New York: McGraw-Hill Book Co 515-534, 2001.
- [14] Toyokuni S: Iron-induced carcinogenesis: the role of redox regulation. Free Radic Biol Med 20: 553-566, 1996.
- [15] Toyokuni S: The origin and future of oxidative stress pathology: From the recognition of carcinogenesis as an iron addiction with ferroptosisresistance to non-thermal plasma therapy. Pathol Int 66: 245-259, 2016.
- [16] Yanatori I, Kishi F: DMT1 and iron transport. Free Radic Biol Med 133: 55-63, 2019.
- [17] Toyokuni S: Role of iron in carcinogenesis: Cancer as a ferrotoxic disease. Cancer Sci 100: 9-16, 2009.
- [18] Toyokuni S, Yanatori I, Kong Y, Zheng H, Motooka Y, et al.: Ferroptosis at the crossroads of infection, aging and cancer. Cancer Sci 111: 2665-2671, 2020.
- [19] Mancias JD, Wang XX, Gygi SP, Harper JW, Kimmelman AC: Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. Nature 509: 105-109, 2014.
- [20] Masaldan S, Clatworthy SAS, Gamell C, Meggyesy PM, Rigopoulos AT, et al.: Iron accumulation in senescent cells is coupled with impaired ferritinophagy and inhibition of ferroptosis. Redox Biol 14: 100-115, 2018.
- [21] Yanatori I, Richardson DR, Toyokuni S, Kishi F: The new role of poly (rC)-binding proteins as iron transport chaperones: Proteins that could couple with inter-organelle interactions to safely traffic iron. Biochim Biophys Acta Gen Subj 1864: 129685, 2020.
- [22] Yanatori I, Richardson DR, Dhekne HS, Toyokuni S, Kishi F: CD63 is Regulated by Iron via the IRE-IRP System and is Important for Ferritin Secretion by Extracellular Vesicles. Blood 2021 doi: 10.1182/blood.2021010995 (in press).
- [23] Mancias JD, Wang X, Gygi SP, Harper JW, Kimmelman AC: Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. Nature 509: 105-109, 2014.
- [24] Yanatori I, Yasui Y, Tabuchi M, Kishi F: Chaperone protein involved in transmembrane transport of iron. Biochem J 462: 25-37, 2014.
- [25] Yanatori I, Richardson DR, Imada K, Kishi F: Iron export through the transporter Ferroportin 1 is modulated by the iron chaperone PCBP2. J Biol Chem 291: 17303-17318, 2016.
- [26] Yanatori I, Richardson DR, Toyokuni S, Kishi F: The iron chaperone poly(rC)-binding protein 2 forms a metabolon with the heme oxygenase 1/cytochrome P450 reductase complex for heme catabolism and iron transfer. J Biol Chem 292: 13205-13229, 2017.
- [27] Shi HF, Bencze KZ, Stemmler TL, Philpott CC: A cytosolic iron chaperone that delivers iron to ferritin. Science 320: 1207-1210, 2008.
- [28] Philpott CC, Jadhav S: The ins and outs of iron: Escorting iron through the mammalian cytosol. Free Radic Biol Med 133: 112-117, 2019.
- [29] Zhang X, Di C, Chen Y, Wang J, Su R, et al.: Multilevel regulation and molecular mechanism of poly (rC)-binding protein 1 in cancer.

FASEB J 34: 15647-15658, 2020.

- [30] Chen C, Lei J, Zheng Q, Tan S, Ding K, et al.: Poly(rC) binding protein 2 (PCBP2) promotes the viability of human gastric cancer cells by regulating CDK2. FEBS Open Bio 8: 764-773, 2018.
- [31] Stockwell BR, Friedmann Angeli JP, Bayir H, Bush AI, Conrad M, et al.: Ferroptosis: A Regulated Cell Death Nexus Linking Metabolism, Redox Biology, and Disease. Cell 171: 273-285, 2017.
- [32] Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, et al.: Ferroptosis: an iron-dependent form of nonapoptotic cell death. Cell 149: 1060-1072, 2012.
- [33] Drakesmith H, Nemeth E, Ganz T: Ironing out Ferroportin. Cell Metab 22: 777-87, 2015.
- [34] Toyokuni S: Novel aspects of oxidative stress-associated carcinogenesis. Antioxid Redox Signal 8: 1373-1377, 2006.
- [35] Toyokuni S: Mysterious link between iron overload and CDKN2A/2B. J Clin Biochem Nutr 48: 46-49, 2011.
- [36] Ito F, Nishiyama T, Shi L, Mori M, Hirayama T, et al.: Contrasting intra- and extracellular distribution of catalytic ferrous iron in ovalbumin-induced peritonitis. Biochem Biophys Res Commun 476: 600-6, 2016.
- [37] Hou W, Xie Y, Song X, Sun X, Lotze MT, et al.: Autophagy promotes ferroptosis by degradation of ferritin. Autophagy 12: 1425-1428, 2016.
- [38] Ito J, Omiya S, Rusu MC, Ueda H, Murakawa T, et al.: Iron derived from autophagy-mediated ferritin degradation induces cardiomyocyte death and heart failure in mice. Elife 10: e62174, 2021.
- [39] Ito F, Yanatori I, Maeda Y, Nimura K, Ito S, et al.: Asbestos conceives Fe(II)-dependent mutagenic stromal milieu through ceaseless macrophage ferroptosis and beta-catenin induction in mesothelium. Redox Biol 36: 101616, 2020.
- [40] Stanton MF, Wrench C: Mechanisms of mesothelioma induction with asbestos and fibrous glass. J Natl Cancer Inst 48: 797-821, 1972.
- [41] Iijima S: Helical Microtubules of Graphitic Carbon. Nature 354: 56-58, 1991.
- [42] Endo M, Strano MS, Ajayan PM, Potential applications of carbon nanotubes, Carbon nanotubes, Springer 2008, pp. 13-62.
- [43] Zhang Q, Huang JQ, Qian WZ, Zhang YY, Wei F: The Road for Nanomaterials Industry: A Review of Carbon Nanotube Production, Post-Treatment, and Bulk Applications for Composites and Energy Storage. Small 9: 1237-1265, 2013.
- [44] Nagai H, Okazaki Y, Chew S, Misawa N, Yamashita Y, et al.: Diameter of multi-walled carbon nanotubes is a critical factor in mesothelial injury and subsequent carcinogenesis. Proc Natl Acad Sci U S A 108: E1330-1338, 2011.
- [45] Nagai H, Okazaki Y, Chew SH, Misawa N, Miyata Y, et al.: Intraperitoneal administration of tangled multiwalled carbon nanotubes of 15 nm in diameter does not induce mesothelial carcinogenesis in rats. Pathol Int 63: 457-462, 2013.
- [46] Wang Y, Okazaki Y, Shi L, Kohda H, Tanaka M, et al.: Role of hemoglobin and transferrin in multi-wall carbon nanotube-induced mesothelial injury and carcinogenesis. Cancer Sci 107: 250-257, 2016.
- [47] Cheng JQ, Jhanwar SC, Klein WM, Bell DW, Lee WC, et al.: p16 alterations and deletion mapping of 9p21-p22 in malignant mesothelioma. Cancer Res 54: 5547-5551, 1994.
- [48] Xio S, Li D, Vijg J, Sugarbaker DJ, Corson JM, et al.: Codeletion of p15 and p16 in primary malignant mesothelioma. Oncogene. 11: 511-515, 1995.
- [49] Jiang L, Akatsuka S, Nagai H, Chew SH, Ohara H, et al.: Iron overload signature in chrysotile-induced malignant mesothelioma. J Pathol 228: 366-377, 2012.
- [50] Akatsuka S, Yamashita Y, Ohara H, Liu YT, Izumiya M, et al.: Fenton reaction induced cancer in wild type rats recapitulates genomic alterations observed in human cancer. PLoS ONE 7: e43403, 2012.
- [51] Okada S: Iron-induced tissue damage and cancer: The role of reactive oxygen free radicals. Pathol Int 46: 311-332, 1996.
- [52] Nakayama M: Macrophage recognition of crystals and nanoparticles. Front Immunol 9: 103, 2018.
- [53] Omori S, Tsugita M, Hoshikawa Y, Morita M, Ito F, et al.: Tim4 recognizes carbon nanotubes and mediates phagocytosis leading to granuloma formation. Cell Rep 34: 108734, 2021.
- [54] Chow A, Schad S, Green MD, Hellmann MD, Allaj V, et al.: Tim-4(+) cavity-resident macrophages impair anti-tumor CD8(+) T cell immunity. Cancer Cell 39: 973-988 e9, 2021.
- [55] Caronni N, Piperno GM, Simoncello F, Romano O, Vodret S, et al.: TIM4 expression by dendritic cells mediates uptake of tumor-associated antigens and anti-tumor responses. Nat Commun 12: 2237, 2021.
- [56] Grosse Y, Loomis D, Guyton KZ, Lauby-Secretan B, El Ghissassi F, et al.: Carcinogenicity of fluoro-edenite, silicon carbide fibres and whiskers, and carbon nanotubes. Lancet Oncol 15: 1427-1428, 2014.
- [57] Sealy C: Carbon nanotubes bring a new touch to robotics. Nano Today 10: 672-673, 2015.
- [58] Pearce CL, Templeman C, Rossing MA, Lee A, Near AM, et al.: Association between endometriosis and risk of histological subtypes of ovarian cancer: a pooled analysis of case-control studies. Lancet Oncol 13: 385-94, 2012.

- [59] Kajiyama H, Suzuki S, Yoshihara M, Tamauchi S, Yoshikawa N, et al.: Endometriosis and cancer. Free Radic Biol Med 133: 186-192, 2018.
- [60] Langseth H, Johansen BV, Nesland JM, Kjaerheim K: Asbestos fibers in ovarian tissue from Norwegian pulp and paper workers. Int J Gynecol Cancer 17: 44-9, 2007.
- [61] Johnson KE, Popratiloff A, Fan Y, McDonald S, Godleski JJ: Analytic comparison of talc in commercially available baby powder and in pelvic tissues resected from ovarian carcinoma patients. Gynecol Oncol 159: 527-533, 2020.
- [62] Slomovitz B, De Haydu C, Taub M, Coleman RL, Monk BJ: Asbestos and ovarian cancer: examining the historical evidence. Int J Gynecol Cancer 31: 122-128, 2021.
- [63] O'Brien KM, Tworoger SS, Harris HR, Anderson GL, Weinberg CR, et al.: Association of powder use in the genital area with risk of ovarian cancer. JAMA 323: 49-59, 2020.
- [64] Johnson KE, Popratiloff A, Fan Y, McDonald S, Godleski JJ: Analytic comparison of talc in commercially available baby powder and in pelvic tissues resected from ovarian carcinoma patients. Gynecol Oncol 159: 527-533, 2020.
- [65] McDonald SA, Fan Y, Welch WR, Cramer DW, Godleski JJ: Migration of Talc From the Perineum to Multiple Pelvic Organ Sites. Am J Clin Pathol 152: 590-607, 2019.
- [66] Goodman JE, Kerper LE, Prueitt RL, Marsh CM: A critical review of talc and ovarian cancer. J Toxicol Environ Health B Crit Rev 23: 183-213, 2020.
- [67] Nagai H, Okazaki Y, Chew SH, Misawa N, Yasui H, et al.: Deferasirox induces mesenchymal-epithelial transition in crocidolite-induced mesothelial carcinogenesis in rats. Cancer Prev Res 6: 1222-30, 2013.
- [68] Ohara Y, Chew SH, Shibata T, Okazaki Y, Yamashita K, et al.: Phlebotomy as a preventive measure for crocidolite-induced mesothelioma in male rats. Cancer Sci 109: 330-339, 2018.
- [69] Shi L, Ito F, Wang Y, Okazaki Y, Tanaka H, et al.: Non-thermal plasma induces a stress response in mesothelioma cells resulting in increased endocytosis, lysosome biogenesis and autophagy. Free Radic Biol Med 108: 904-917, 2017.
- [70] Jiang L, Zheng H, Lyu Q, Hayashi S, Sato K, et al.: Lysosomal nitric oxide determines transition from autophagy to ferroptosis after exposure to plasma-activated Ringer's lactate. Redox Biol 43: 101989, 2021.
- [71] Ebina Y, Okada S, Hamazaki S, Ogino F, Li JL, et al.: Nephrotoxicity and renal cell carcinoma after use of iron- and aluminum- nitrilotriacetate complexes in rats. J Natl Cancer Inst 76: 107-113, 1986.
- [72] Li JL, Okada S, Hamazaki S, Ebina Y, Midorikawa O: Subacute nephrotoxicity and induction of renal cell carcinoma in mice treated with ferric nitrilotriacetate. Cancer Res 47: 1867-1869, 1987.
- [73] Nagai H, Ishihara T, Lee WH, Ohara H, Okazaki Y, et al.: Asbestos surface provides a niche for oxidative modification. Cancer Sci 102: 2118-2125, 2011.